

# Community baseline vertebrate biodiversity surveys in the Hornsby Council region using eDNA - Spring 2023

## Wednesday, 20 December 2023

Project number:	ED_2406CR1
Client:	Hornsby Shire Council
Prepared for:	Petra Holland, David Bolton
Report prepared by:	Emma Walker, Dr Reid Tingley
Report approved by:	Sarah Hale
Lab analyst:	Dr Rachael Impey
Assay(s):	<i>Vertebrate (12S)</i>
Filter used:	1.2 µm EnviroDNA manual syringe disc filter

## Highlights

- At each of the 41 sites, 2 water samples were collected.
- 67% of taxa were resolved at the species level.
- Across all sites, 97 taxa were detected, including 1 threatened species (*Pteropus poliocephalus*).
- Taxon richness at the site level ranged from 3 to 29.
- *Anguilla reinhardtii* was the most commonly detected taxon.
- >5000 reads were obtained for 96% of metabarcoding samples.

## Background

Environmental DNA (eDNA) methods are being used routinely to monitor aquatic animals including fish, amphibians and mammals across waterways, estuaries and wetlands throughout Australian catchments. Here we use a vertebrate eDNA metabarcoding assay to screen 82 eDNA samples taken from 41 sites throughout the Hornsby council region, New South Wales to provide a baseline biodiversity assessment of vertebrate species during Spring, 2023.

## Methods

### Sampling

During Spring 2023, 82 water samples were collected from 41 sites by Hornsby council staff and citizen scientists. At each site, 2 replicate samples were collected by passing up to 2,000 mL of water (mean = 907 mL) through a 1.2 µm EnviroDNA manual syringe disc filter. Filtration was undertaken on-site to reduce DNA degradation during transport of water samples. Filters were stored out of sunlight and at ambient temperature before being transported to the laboratory for processing.

### Analysis

DNA was extracted from filters using a Qiagen PowerSoil Kit that minimises compounds that can inhibit PCR reactions in environmental samples. Library construction involved two rounds of PCR, whereby the first round employed gene-specific primers to amplify the target region and the second round incorporated sequencing adapters and unique barcodes for each sample-amplicon combination included in the library. Negative controls were included during library construction. Negative controls consisted of the extraction negative as well as PCR negatives, in which nuclease-free water was used in place of DNA during both rounds of PCR. Sequencing was carried out on an Illumina sequencing platform.

Following quality control filtering to remove primer sequences, truncated reads, and low-frequency reads, DNA sequences were clustered into Operational Taxonomic Units (OTUs) on the basis of sequence similarity. Taxonomic assignment was performed with VSEARCH software (Rognes et al. 2016), whereby each OTU cluster was assigned a species identity using a threshold of 95% by comparing against a reference sequence database. Where a species could not be assigned (i.e., reference database was deficient and/or taxa were poorly-characterised), taxonomic assignments were manually vetted by first obtaining a list of possible species through BLASTN searches against the public repository Genbank ([www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)), followed by elimination of species on the basis of their geographic distributions, using information from the Atlas of Living Australia and other relevant data sources. In cases where an OTU could not be adequately resolved to a single species (e.g., due to shared haplotypes), either a list of multiple species is included, or the OTU is assigned to the lowest taxonomic rank without further classification.

## Results

A total of 82 samples were analysed from 41 sites across the Hornsby Council region, NSW using a 1.2 µm EnviroDNA manual syringe disc filter. Raw data on per-sample detections can be found in accompanying spreadsheet (ED\_2406CR1\_Hornsby\_Vert\_Data). The spreadsheet provides the taxa detected in each sample, as well as the number of sequence reads for each taxon. Reads should not be directly interpreted as taxa abundance. While some studies have shown a positive correlation between read numbers and abundance, reads can also be influenced by a number of other variables. Reads may be used to help assign a level of confidence in species detection along with the number of replicates in which the species was detected.

Overall, 97 vert taxa were detected, including 12 introduced species and 1 species listed at the Federal and/or State level; Grey-headed flying-fox (*Pteropus poliocephalus*) (listing data from <https://www.environment.gov.au/sprat-public/action/report>). Six frog, 25 fish, 6 reptile, 36 bird, and 24 mammal taxa were detected. The number of vertebrate taxa at each site (across all replicate samples) ranged from 3 to 29. The number of native taxa per site varied from 1 to 16 (Figure 1).

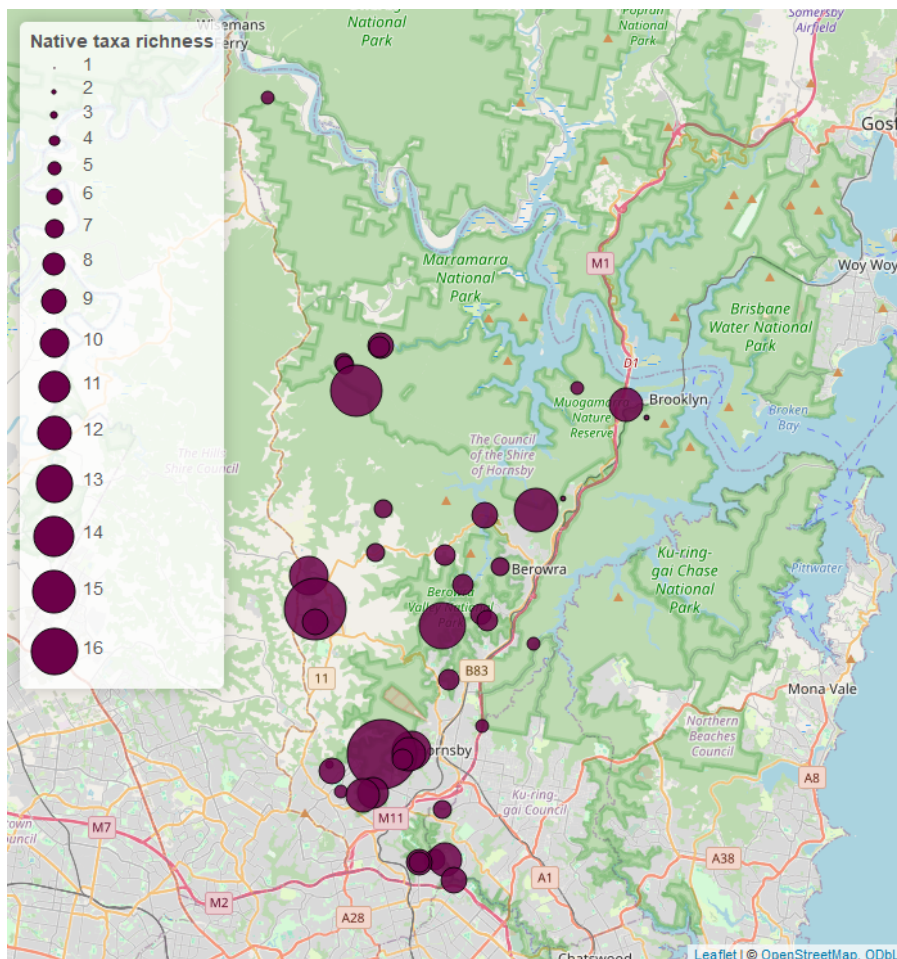


Figure 1. Native vertebrate species richness at the 41 sampled sites. Marker size is proportional to detected species richness. Note that mapped native richness only includes taxa resolved at the species level.

Most taxa were resolved at the species level (67% of all taxa). The fact that some taxa could not be resolved at the species level is likely due to inadequate genetic sequence data available in the reference library for the region. Further reference sequences for species that are not currently captured in the reference database are needed to fully evaluate the potential for the 12S region to resolve these taxa to a species or genus level. Unresolved taxa can also arise due to limitations with the target region (e.g., 12S, 16S) and metabarcoding assays in general, whereby only a very small subset of the entire genome is interrogated for the purpose of species identification. Consequently, there is not always enough genetic variation in that short marker sequence to definitively assign it to a species.

A summary of the frequency of occurrence of each vertebrate species across all samples and sites is provided in Table 1.

Table 1. Number of detections and number of occupied sites for each vertebrate taxon.

Group	Taxa	Common name	N detections	N sites
Birds	<i>Acanthagenys rufogularis</i>	Spiny-cheeked honeyeater	1	1
	<i>Acanthiza pusilla</i>	Brown thornbill	2	2
	<i>Alectura lathamii</i>	Australian brush-turkey	3	3
	<i>Alisterus scapularis</i>	Australian king-parrot	3	3
	<i>Anatidae</i>	Family of waterbirds that includes ducks, geese and swans	6	5
	<i>Cacatua</i>	Genus of cockatoo	1	1
	<i>Cacatua galerita</i>	Sulphur-crested cockatoo	13	10
	<i>Charadriiformes</i>	Order of shorebirds	1	1
	<i>Chenonetta jubata</i>	Australian wood duck	1	1
	<i>Columba livia</i>	Domestic pigeon	3	3
	<i>Cormobates leucophaea</i>	White-throated treecreeper	2	2
	<i>Corvus</i>	Genus of crows and ravens	1	1
	<i>Dacelo novaeguineae</i>	Laughing kookaburra	6	4
	<i>Eolophus roseicapilla</i>	Galah	2	2
	<i>Gallinula tenebrosa</i>	Dusky moorhen	6	4
	<i>Malurus lamberti</i>	Variiegated fairy-wren	1	1
	<i>Manorina</i>	Genus of Australian honeyeaters and miners	9	6
	<i>Meliphaga lewinii</i>	Lewin's honeyeater	1	1
	<i>Meliphagidae</i>	Family of honeyeaters	4	3
	<i>Menura</i>	Superb lyrebird	1	1
	<i>novaehollandiae</i>			
	<i>Microcarbo melanoleucos</i>	Little pied cormorant	5	4

Group	Taxa	Common name	N detections	N sites
	<i>Ocyphaps lophotes</i>	Crested pigeon	1	1
	<i>Pachycephala</i>	Genus of whistlers	1	1
	<i>Passeriformes</i>	Order of perching birds	8	7
	<i>Phalacrocorax sulcirostris</i>	Little black cormorant	4	3
	<i>Platycercus eximius</i>	Eastern rosella	9	8
	<i>Podargus strigoides</i>	Tawny frogmouth	2	2
	<i>Poodytes</i>	Genus of grassbirds	2	2
	<i>Ptilonorhynchus violaceus</i>	Satin bowerbird	1	1
	<i>Sericornis frontalis</i>	White-browed scrubwren	1	1
	<i>Sturnidae</i>	Family of starlings	1	1
	<i>Sturnus vulgaris</i>	Common starling	1	1
	<i>Trichoglossus</i>	Genus of lorikeet	6	6
	<i>Turdus</i>	Genus of thrush	4	3
	<i>Turdus philomelos</i>	Song thrush	2	2
	<i>Zanda</i>	Genus of black cockatoo	2	1
	<i>Actinopteri</i>	Class of fish, unassigned	14	9
	<i>Anguilla</i>	Genus of freshwater eels	34	21
	<i>Anguilla reinhardtii</i>	Longfin eel	60	35
	<i>Carassius auratus</i>	Goldfish	3	3
	<i>Clupeidae</i>	Family of herrings and sprats	1	1
	<i>Cyprinus carpio</i>	European carp	7	4
	<i>Galaxias</i>	Genus of galaxiids	17	11
	<i>Gambusia</i>	Genus of mosquitofish	10	6
	<i>Gobiomorphus australis</i>	Striped gudgeon	24	13
	<i>Gobiomorphus coxii</i>	Cox's gudgeon	48	30
	<i>Gracilimugil argenteus</i>	Flat-tail mullet	2	1
Fishes & eels	<i>Hypseleotris</i>	Genus of carp gudgeons	5	3
	<i>Melanotaenia</i>	Genus of rainbowfish	1	1
	<i>Mugil cephalus</i>	Sea mullet	2	1
	<i>Mugilidae</i>	Family of mullet	1	1
	<i>Mugilogobius platynotus</i>	Flatback mangrovegoby	1	1
	<i>Notesthes robusta</i>	Bullrout	2	1
	<i>Percalates novemaculeata</i>	Australian bass	14	8
	<i>Philypnodon grandiceps</i>	Flatheaded gudgeon	7	4
	<i>Philypnodon macrostomus</i>	Dwarf flathead gudgeon	2	1
	<i>Retropinna semoni</i>	Australian smelt	10	6

Group	Taxa	Common name	N detections	N sites
	<i>Salmo</i>	Genus of salmon and trout	1	1
	<i>Scomber</i>	Genus of mackerels	1	1
	<i>Tandanus tandanus</i>	Eel-tailed catfish, freshwater catfish	2	2
	<i>Tetractenos</i>	Genus of toadfish	2	1
Frogs	<i>Crinia signifera</i>	Common froglet	34	22
	<i>Limnodynastes peronii</i>	Striped marsh frog	23	14
	<i>Litoria fallax</i>	Eastern dwarf tree frog	3	2
	<i>Litoria gracilentata</i>	Dainty green tree frog	1	1
	<i>Litoria peronii</i>	Peron's tree frog	12	10
	<i>Litoria phyllochroa</i>	Leaf green tree frog	40	25
	<i>Bos taurus</i>	Cattle	8	7
	<i>Canis lupus</i>	Dog/dingo	18	14
	<i>Diprotodontia</i>	Order of marsupials that includes kangaroos, wallabies, possums	15	12
	<i>Eptesicus vulturinus</i> or <i>Vespadelus vulturinus</i>	Little forest bat	1	1
	<i>Hydromys chrysogaster</i>	Rakali	3	2
	<i>Macropodidae</i>	Family of marsupials that includes kangaroos, wallabies	3	2
	<i>Macropus giganteus</i>	Eastern grey kangaroo	1	1
	<i>Mormopterus planiceps</i>	Southeastern free-tailed bat	1	1
	<i>Mus musculus</i>	House mouse	4	4
	<i>Myotis adversus</i>	Large foot bat	1	1
Mammals	<i>Notamacropus</i>	Genus of wallaby	1	1
	<i>Oryctolagus cuniculus</i>	European rabbit	2	2
	<i>Ovis aries</i>	Sheep	4	3
	<i>Perameles</i>	Genus of bandicoots	1	1
	<i>Petaurus breviceps</i>	Sugar glider	2	2
	<i>Pseudocheiridae</i>	Family of ring-tailed possums	5	4
	<i>Pseudocheirus peregrinus</i>	Eastern ring-tailed possum	1	1
	<i>Pteropus poliocephalus</i>	Grey-headed flying-fox	9	7
	<i>Rattus</i>	Genus of rodents	5	5
	<i>Rattus fuscipes</i>	Bush rat	5	4
	<i>Rattus norvegicus</i>	Brown rat	3	2
	<i>Rattus rattus</i>	Black rat	9	8
	<i>Sus scrofa</i>	Pig	11	9
	<i>Trichosurus vulpecula</i>	Common brush-tailed possum	6	6
Reptiles	<i>Chelodina</i>	Genus of snake necked turtles	1	1

Group	Taxa	Common name	N detections	N sites
	<i>Concinnia tenuis</i>	Barred-sided skink	3	2
	<i>Eulamprus quoyii</i>	Eastern water skink	8	6
	<i>Myuchelys latisternum</i>	Saw-shelled turtle	1	1
	<i>Saproscincus mustelinus</i>	Weasel skink	4	3
	<i>Scincidae</i>	Genus of skinks	2	1

Figure 3, below, shows similar data to those presented in the table above. Rather than focusing on the number of detections, however, this figure shows the percentage of reads assigned to each taxon.

### Quality Control

- Amplification success was confirmed by gel electrophoresis.
- The following controls were used:
  - o 2 extraction controls
  - o 4 mock communities
- The total number of reads was 4,884,394.
- The median number of reads per sample was 59,458.5 (range = 0 – 92,983).
- Out of 82 samples analysed, 3 samples were labelled dropouts (fewer than 5,000 non-human reads).
- Numbers of reads in negative controls were below the acceptable threshold.
- All mock community positive controls produced reads of expected species, with no contamination from other species.



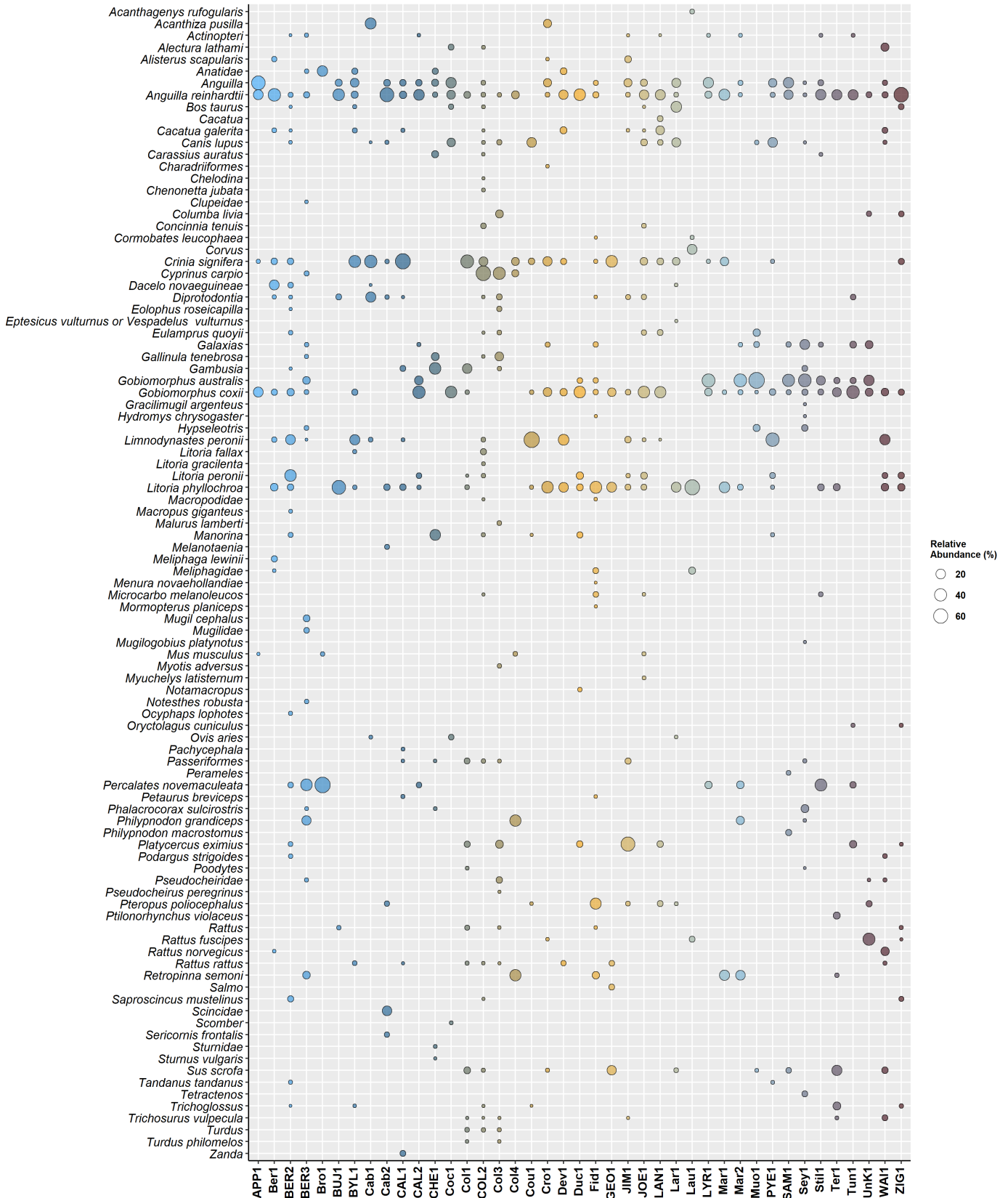


Figure 3. Percentage of reads assigned to each vertebrate taxon.



## References

Rognes T, Flouri T, Nichols B, Quince C, Mahé F. VSEARCH: a versatile open source tool for metagenomics. PeerJ. 2016 Oct 18;4:e2584. doi: 10.7717/peerj.2584. PMID: 27781170; PMCID: PMC5075697.

### Disclaimer

The professional analysis and advice in this report has been prepared for the exclusive use of the party or parties to whom it is addressed (the addressee) and for the purposes specified in it. This report is supplied in good faith and reflects the knowledge, expertise and experience of the consultants involved. The report must not be published, quoted or disseminated to any other party without prior written consent from EnviroDNA Pty Ltd.

EnviroDNA Pty Ltd accepts no responsibility whatsoever for any loss occasioned by any person acting or refraining from action as a result of reliance on the report. In conducting the analysis in this report EnviroDNA Pty Ltd has endeavoured to use what it considers is the best information available at the date of publication including information supplied by the addressee. Unless stated otherwise EnviroDNA Pty Ltd does not warrant the accuracy of any forecast or prediction in this report.